

# A confidence framework for fish detection through environmental DNA metabarcoding:

## Achieving confidence in absence

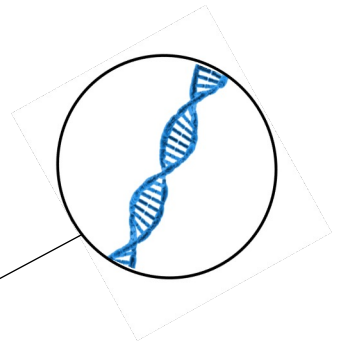
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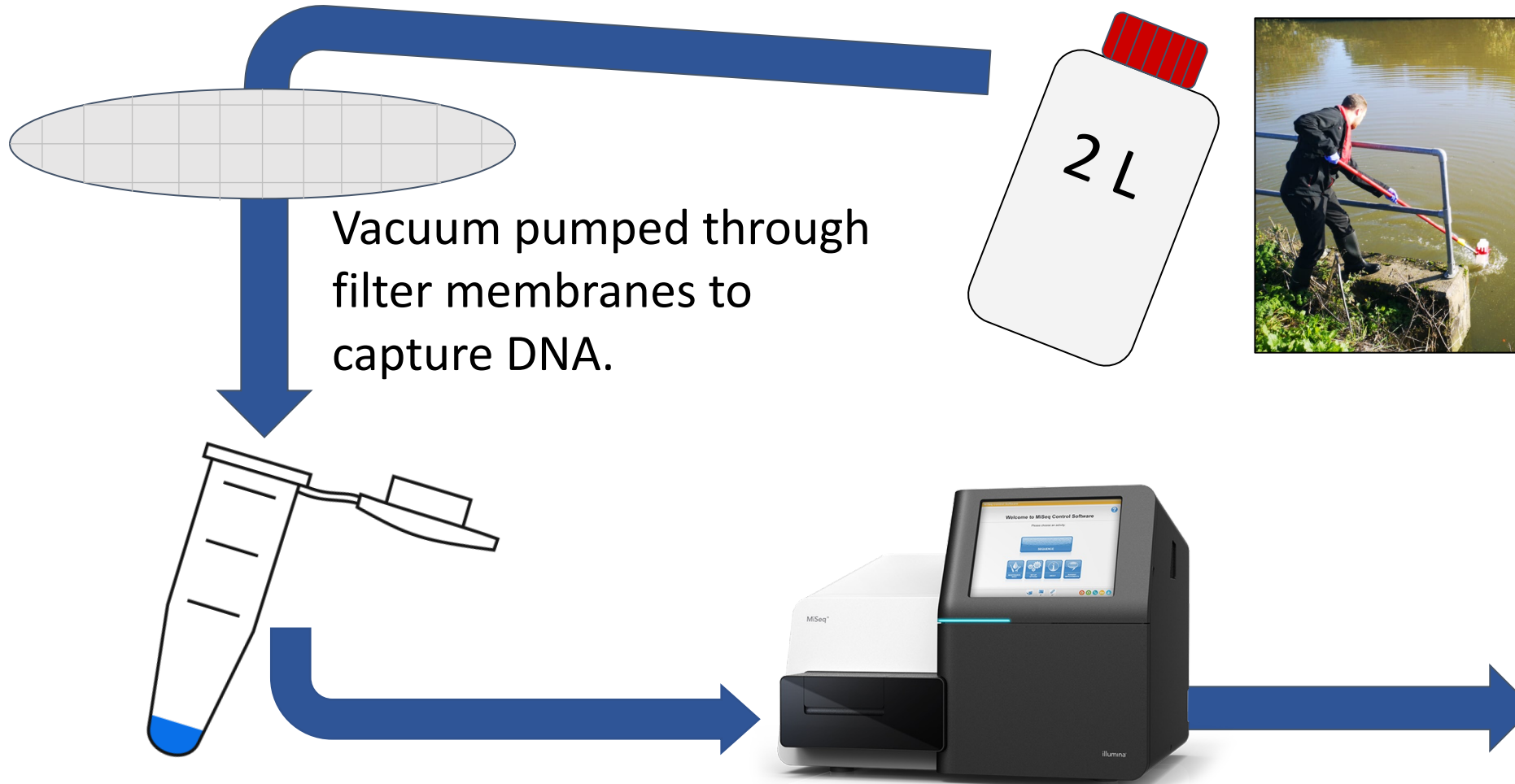


# Environmental DNA

- As species interact with the environment, DNA is shed.
- This DNA is referred to as Environmental DNA (eDNA).
- eDNA can be sampled, to identify species present in the watercourse.



# eDNA metabarcoding workflow



Water samples are obtained from the field.



```
>yneN
TTAATGCCTCTTCTCATTCTCTCTGCTGTCCGACAGCAGAGAAGAAATTCCTCATTGAC
TATTAATTCGCAATTTGGCTCACATGGATTAATTAACACATACTATAAGATATAAACT
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TCTCCAGCCTGTTAATAAGCATACTGATAACGATTTTAACTGTTATCCGCTAAATAA
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A
>yegR
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GGGAAGGTGAGATGAAAAAGATAGCTGATATATCATTAAATAGTATTTTATATGTCG
G
>emrK
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T
>evgA
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CTGATTTCTCATTTCATGCTCACCCAATATGATGGCGGCTTTTCAAACTGTTAAAGA
ATCAGGTAAGTATGAAACGTTAATATGACCAGATGTTGACAGCAATTCCTGGCATCT
C
```

Sequenced reads are assigned to species.

Sellers *et al.* 2018

# Aims

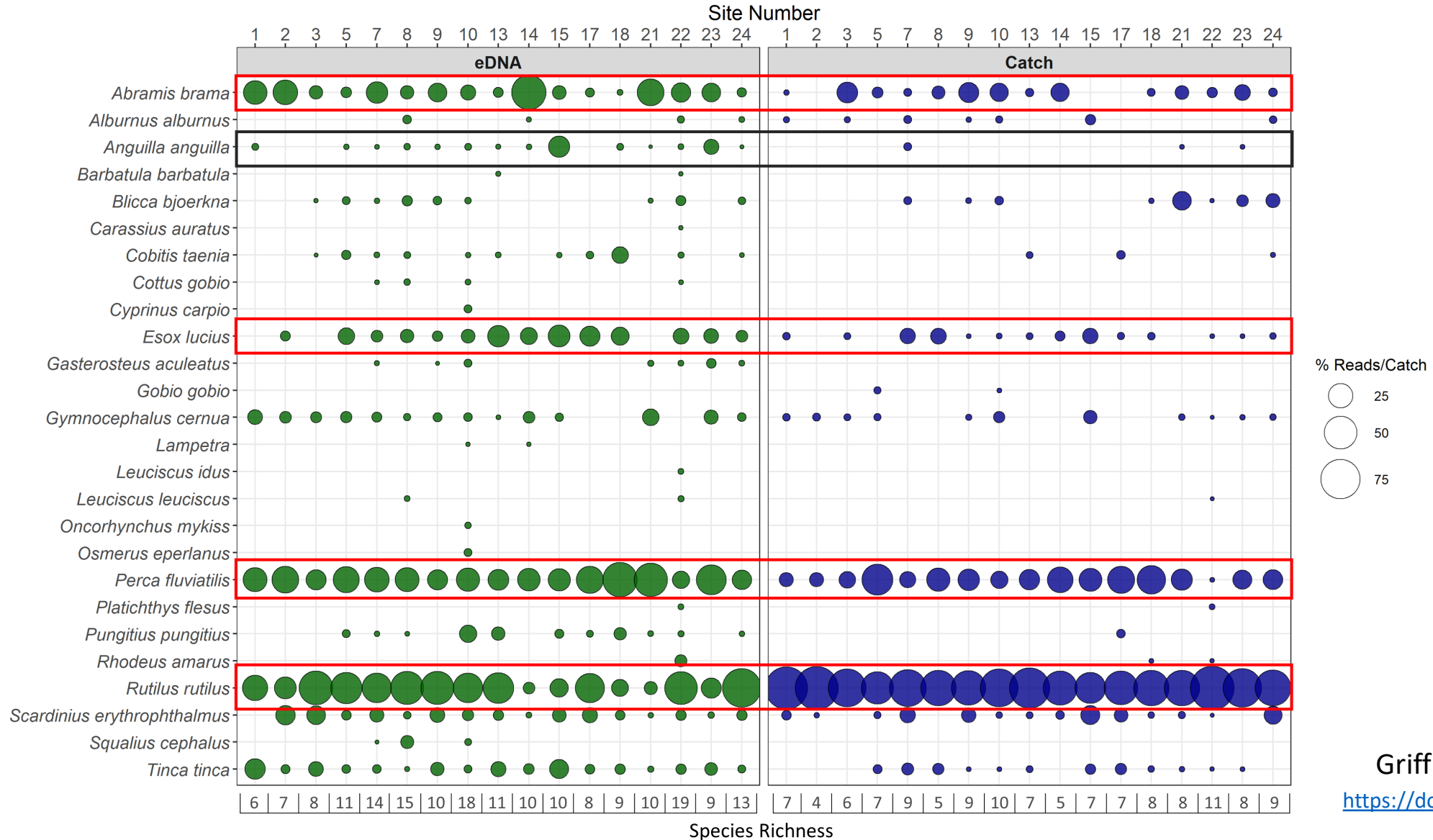
- Optimise eDNA methods to determine species composition in heavily managed catchments.
- Apply this method as a tool to enable targeted management.

**End goal**

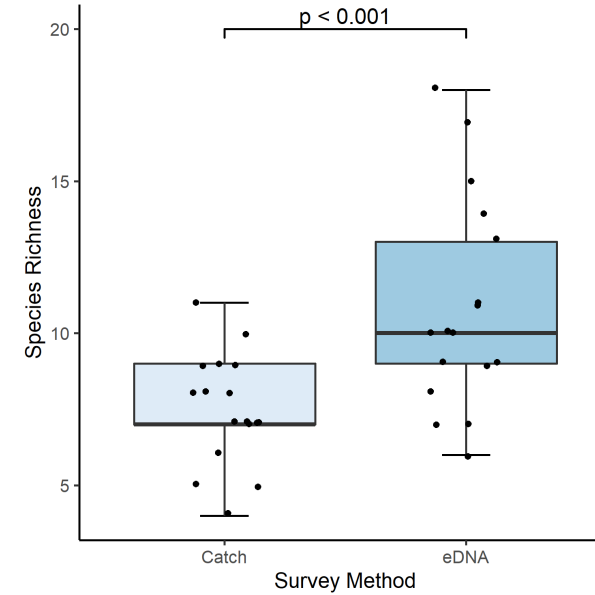
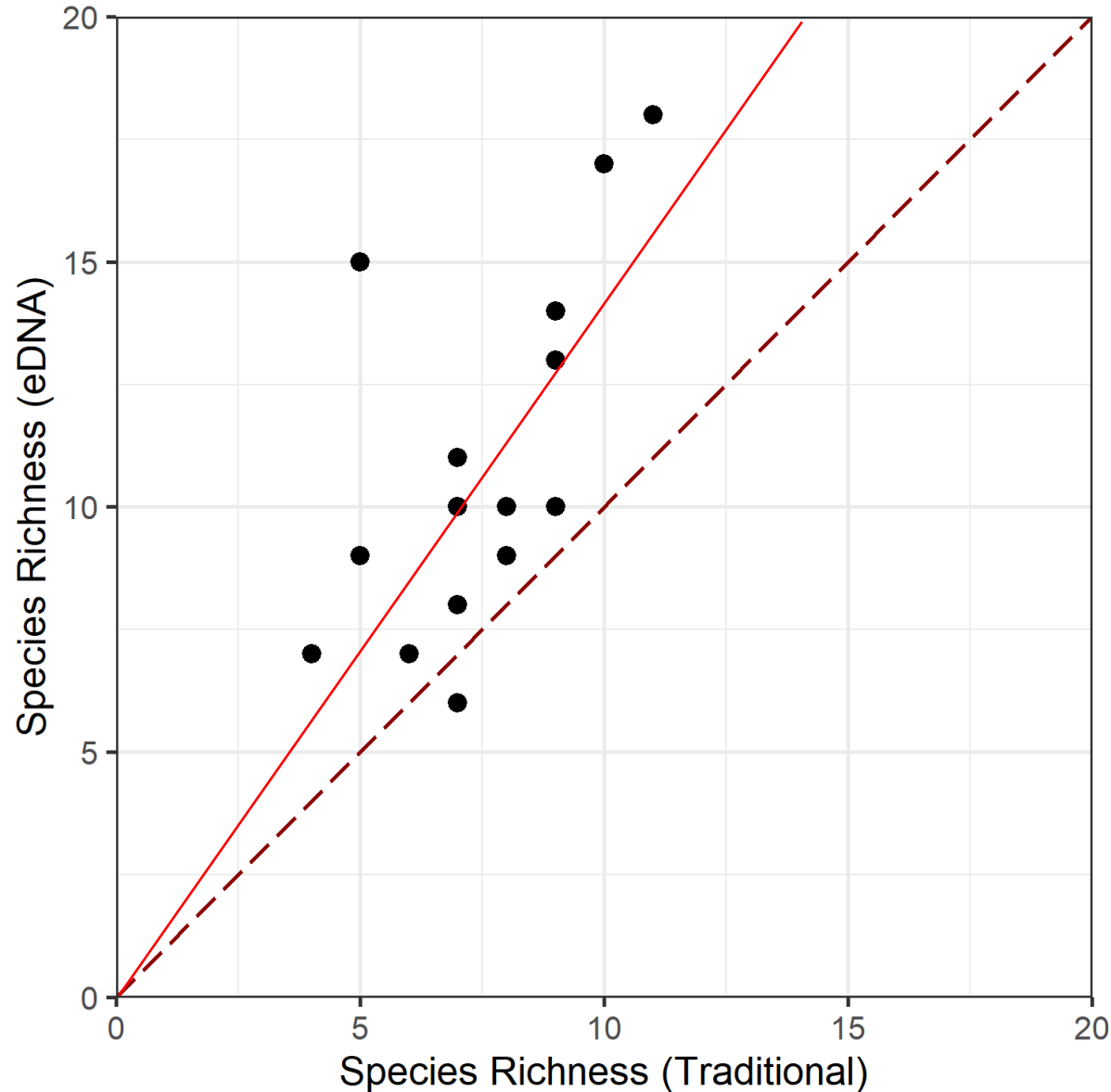


**Integrate eDNA based monitoring  
into prioritisation frameworks**

# This method has already proven effective in highly managed catchments in the UK

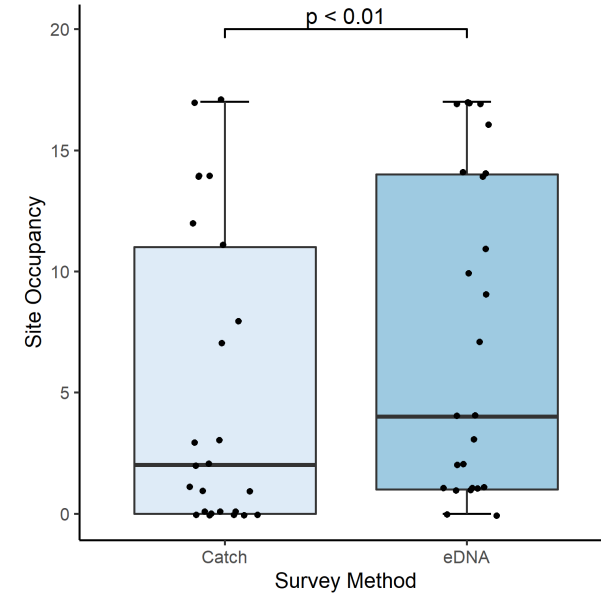
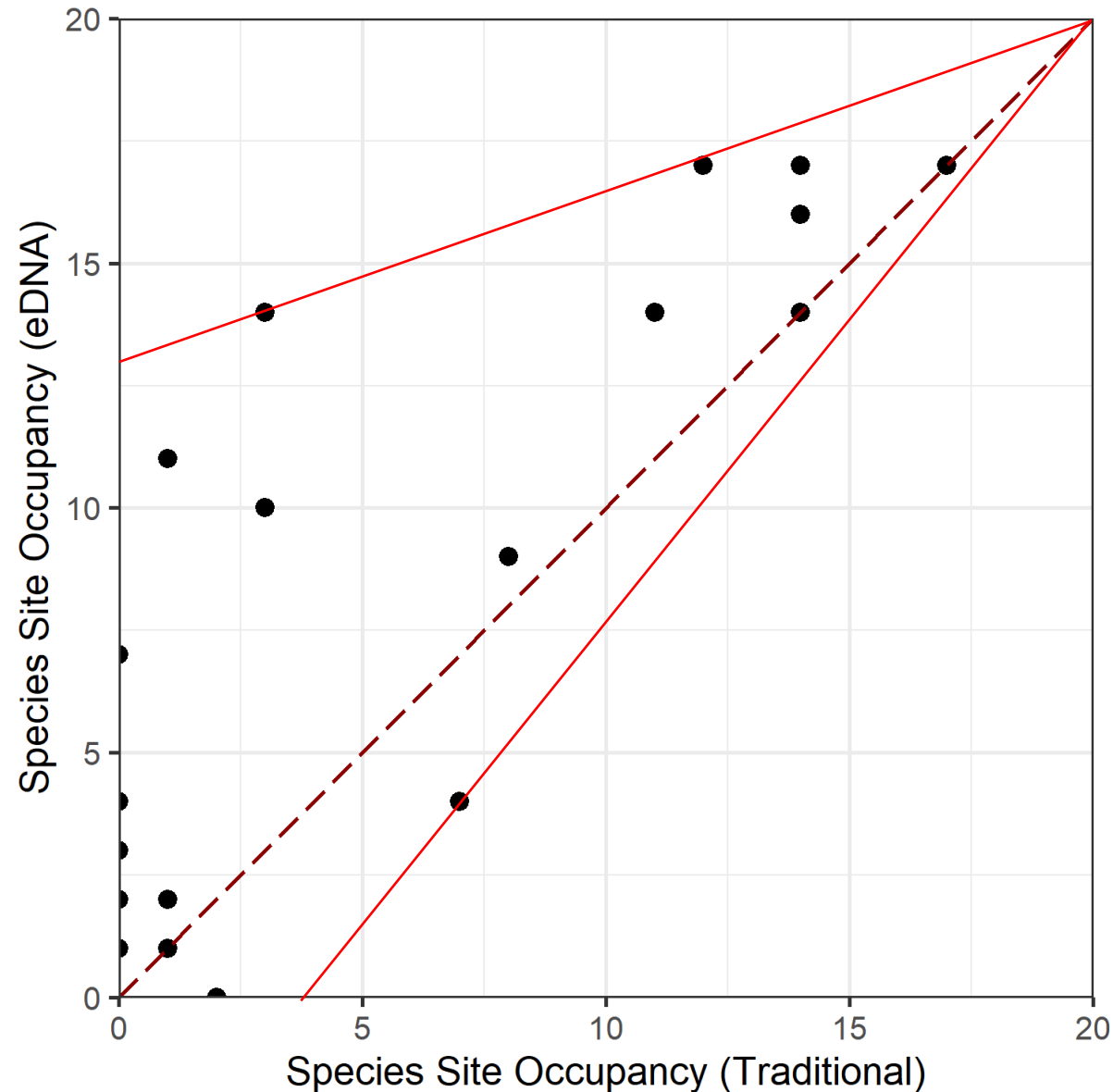


# Species Richness



- Species richness was higher for 16/17 sites when using eDNA.
- Average species richness across the catchment was significantly higher when using eDNA methods.

# Species Site Occupancy



- eDNA site occupancy was  $\geq$  traditional methods for 22/25 fish species detected across all surveys.
- Catchment wide occupancy was significantly higher when using eDNA methods.

# Prioritising fish pass solutions

- There are over 900 water pumping stations in England alone.
- Lots of innovative solutions to fish passage in development.
- We need informed prioritisation to make use of limited resources!





# Priority species

- Specific legislation means some species take high priority.
- For these, a ‘false negative’ would be in breach of policy.
- To enable targeted management, we must be **confident** of where priority species are **present**, and **absent**.

COUNCIL REGULATION (EC) No 1100/2007  
of 18 September 2007  
establishing measures for the recovery of the stock of European eel

2009 No. 3344  
FISHERIES, ENGLAND AND WALES  
RIVER, ENGLAND AND WALES  
The Eels (England and Wales) Regulations 2009



# Aims

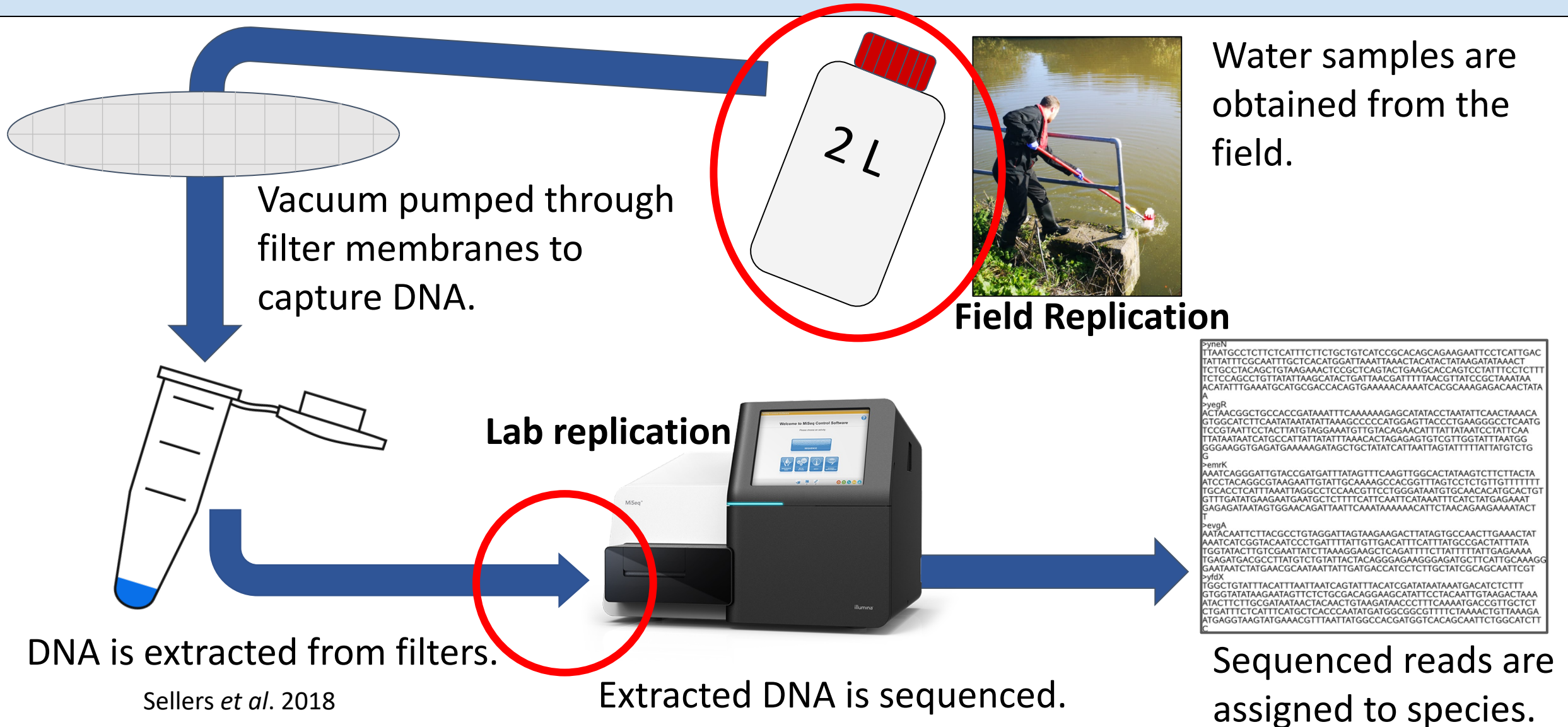
- Determine the sensitivity of our eDNA metabarcoding workflow.
- Optimise the number of sample and lab replicates required.

**End goal**



**Sampling designs tailor-made to  
suit end-user requirements.**

# eDNA metabarcoding workflow



Sellers *et al.* 2018

# Levels of replication

## Site

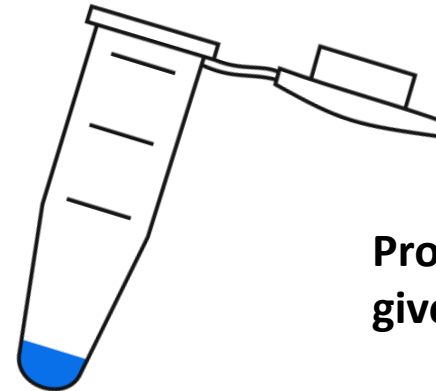


## Field replicate:



Probability of DNA in sample, given presence at site?

## Lab replicates:



Probability DNA detection, given presence in sample?

# Pizza analogy

(a) Site:



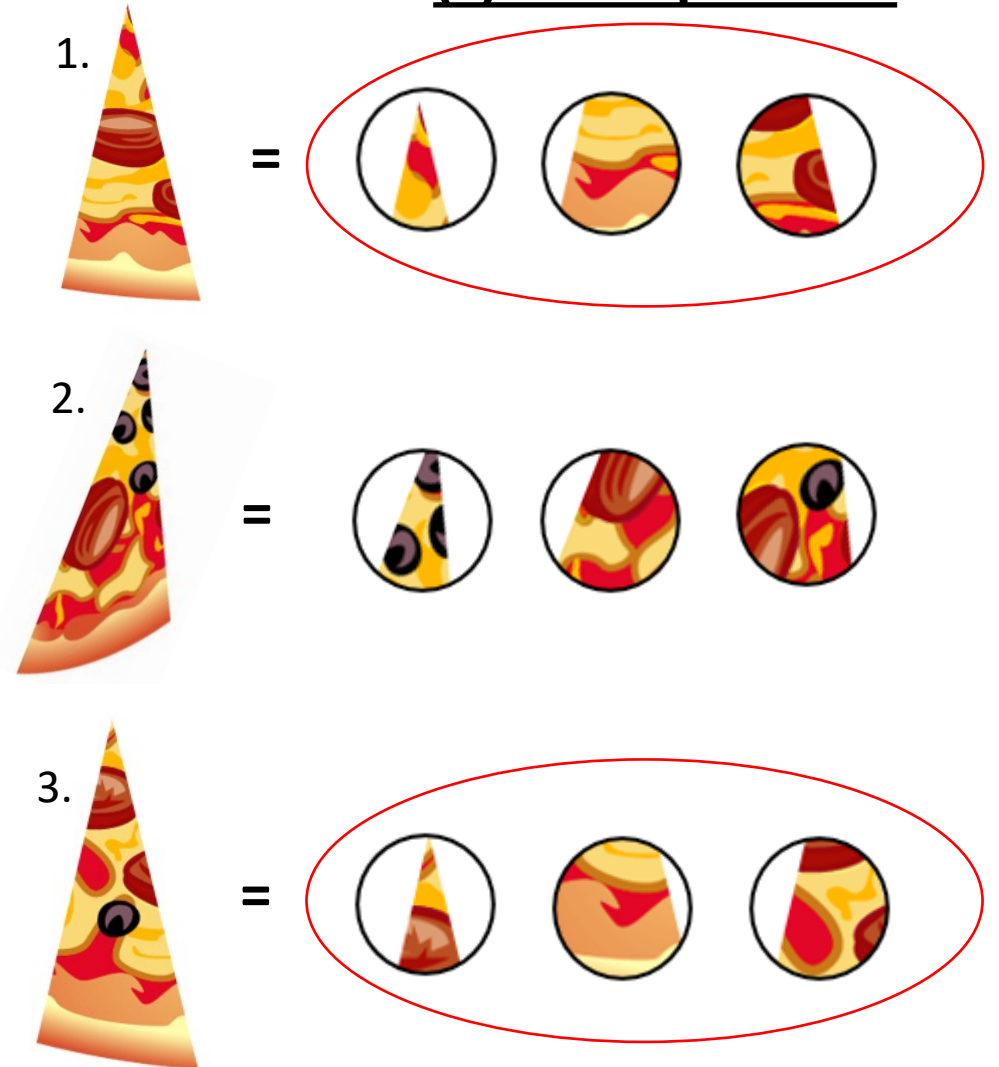
(b) Samples:



No olives?  
(false negative)



(c) Lab replicates:



# Pizza analogy

(a) Site:

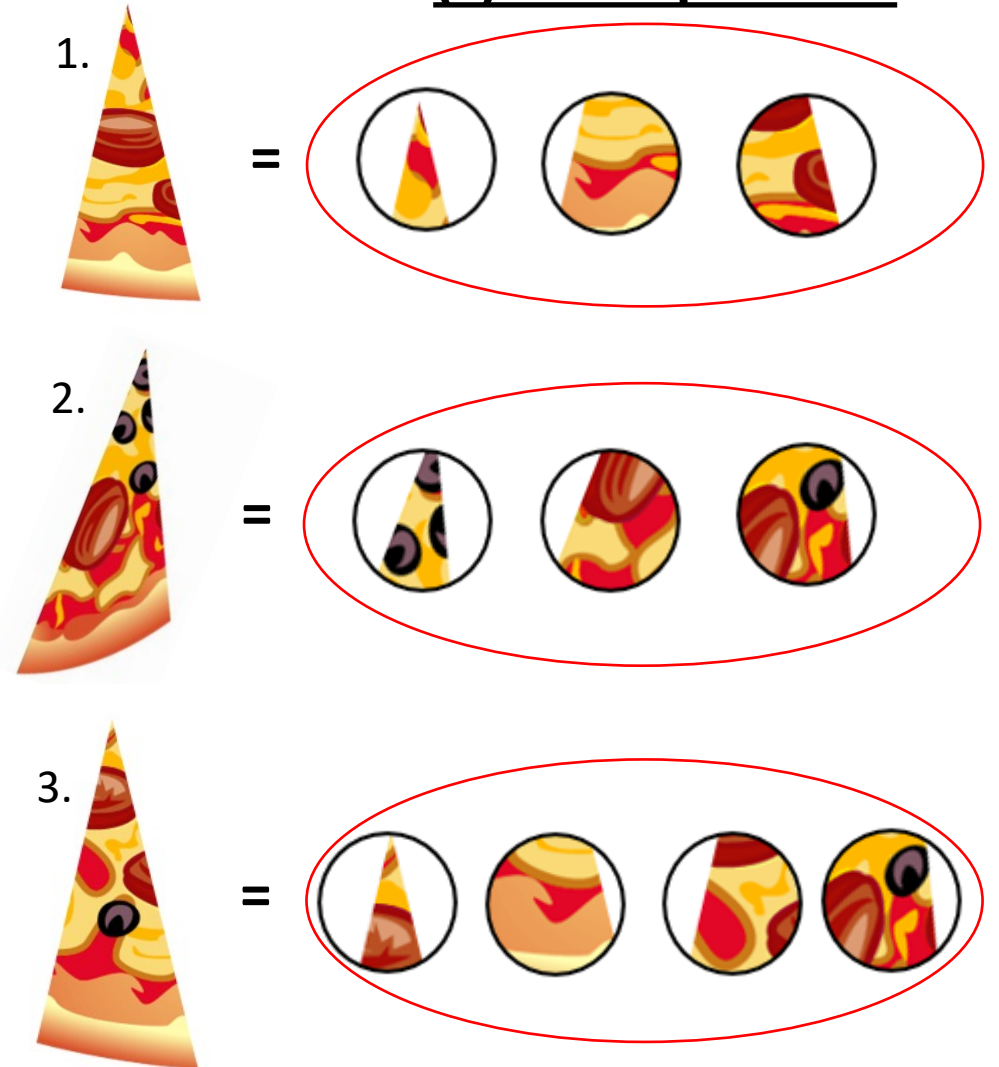


olives ✓

(b) Samples:

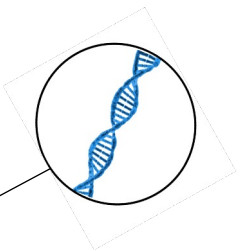


(c) Lab replicates:



# Our Study

- **Field replication** – 44 sites each with 10x samples processed
- **Lab replication** – 10x samples obtained at a site each underwent 10x PCR replicates



# Results

- We confirmed eel presence at 17 / 44 of our study sites.
- But... does this mean we can be confident the remaining 27 should be classified as **absent** for eels?





# Our model

a = probability of occupancy at a site

b = conditional probability of DNA presence in a sample given occupancy at the site

c = conditional probability of DNA detection in a replicate given presence in the sample

If  $n$  samples are taken, with  $m$  PCR replicates, and no eels are detected, then the probability of absence is:

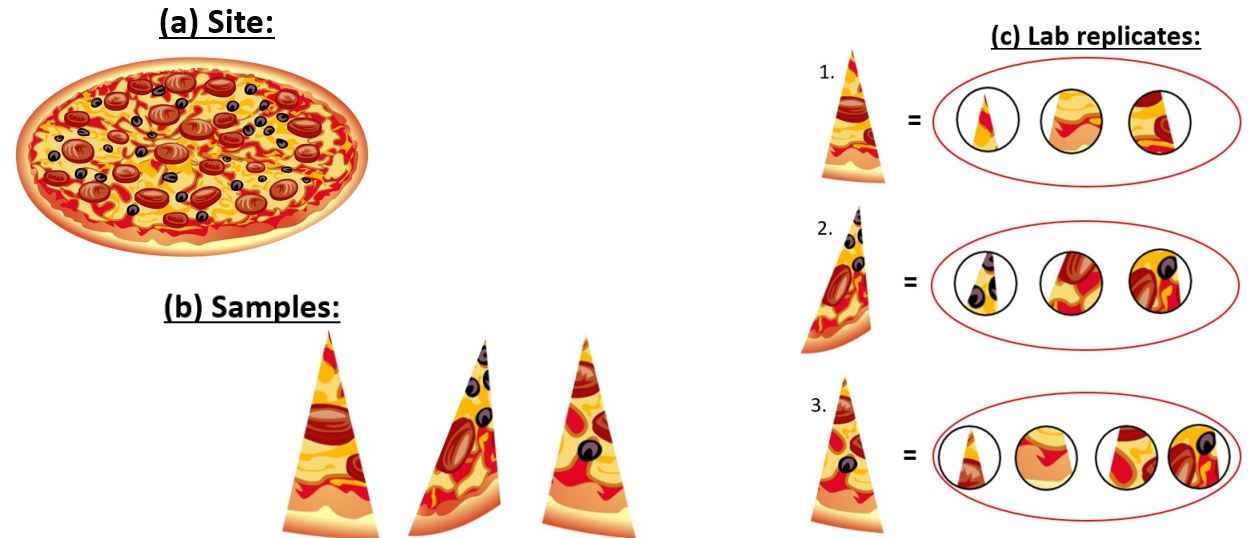
$$(1-a)/(1-a+a*(1-b+b*(1-c)^m)^n)$$

When we apply this to eels:

a = 38.7%

b = 86.8%

c = 25.7%



# Confidence of absence for Eels

	1 sample	2 samples	3 samples	4 samples	5 samples	6 samples	7 samples	8 samples	9 samples	10 samples
1 replicate	0.67	0.72	0.77	0.81	0.85	0.88	0.90	0.92	0.94	0.95
2 replicates	0.72	0.81	0.87	0.92	0.95	0.97	0.98	0.99	0.99	1.00
3 replicates	0.76	0.87	0.93	0.97	0.98	0.99	1.00	1.00	1.00	1.00
4 replicates	0.80	0.91	0.96	0.98	0.99	1.00	1.00	1.00	1.00	1.00
5 replicates	0.83	0.94	0.98	0.99	1.00	1.00	1.00	1.00	1.00	1.00
6 replicates	0.85	0.95	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00
7 replicates	0.87	0.96	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00
8 replicates	0.88	0.97	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00
9 replicates	0.89	0.98	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
10 replicates	0.90	0.98	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
11 replicates	0.91	0.98	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
12 replicates	0.91	0.98	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
13 replicates	0.91	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
14 replicates	0.92	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
15 replicates	0.92	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
16 replicates	0.92	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
17 replicates	0.92	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
18 replicates	0.92	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
19 replicates	0.92	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
20 replicates	0.92	0.99	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00

**Optimal strategy**

**Wasted effort**

a = 38.7%

b = 86.8%

c = 25.7%

<95% confidence

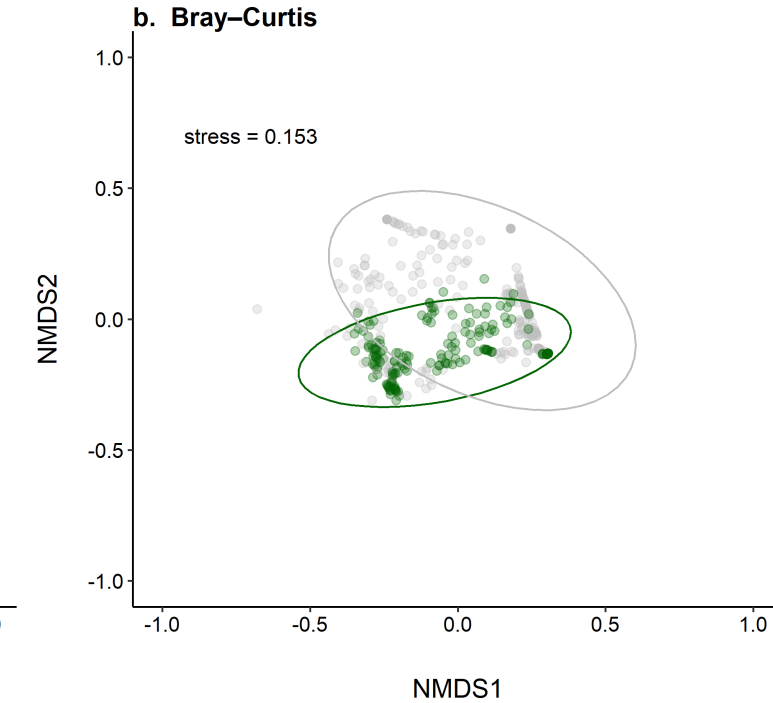
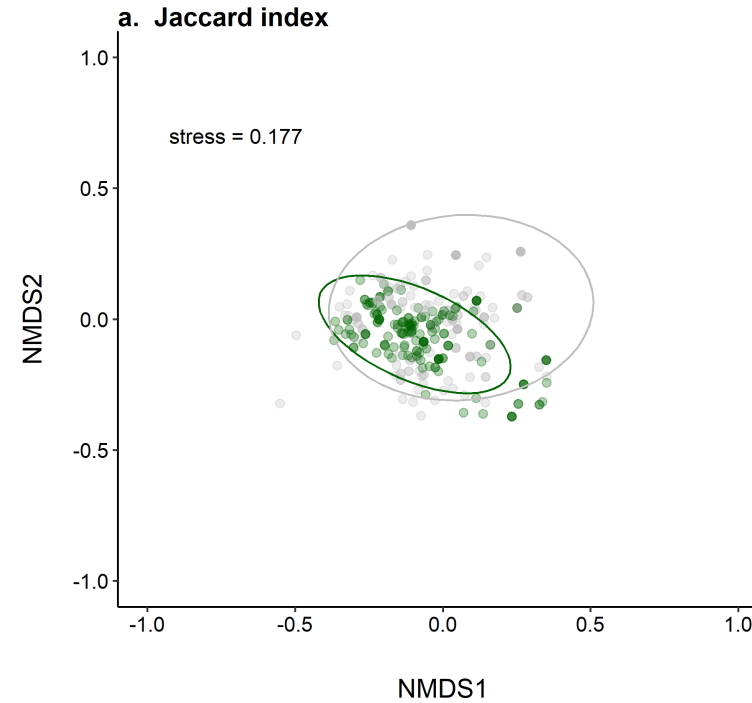
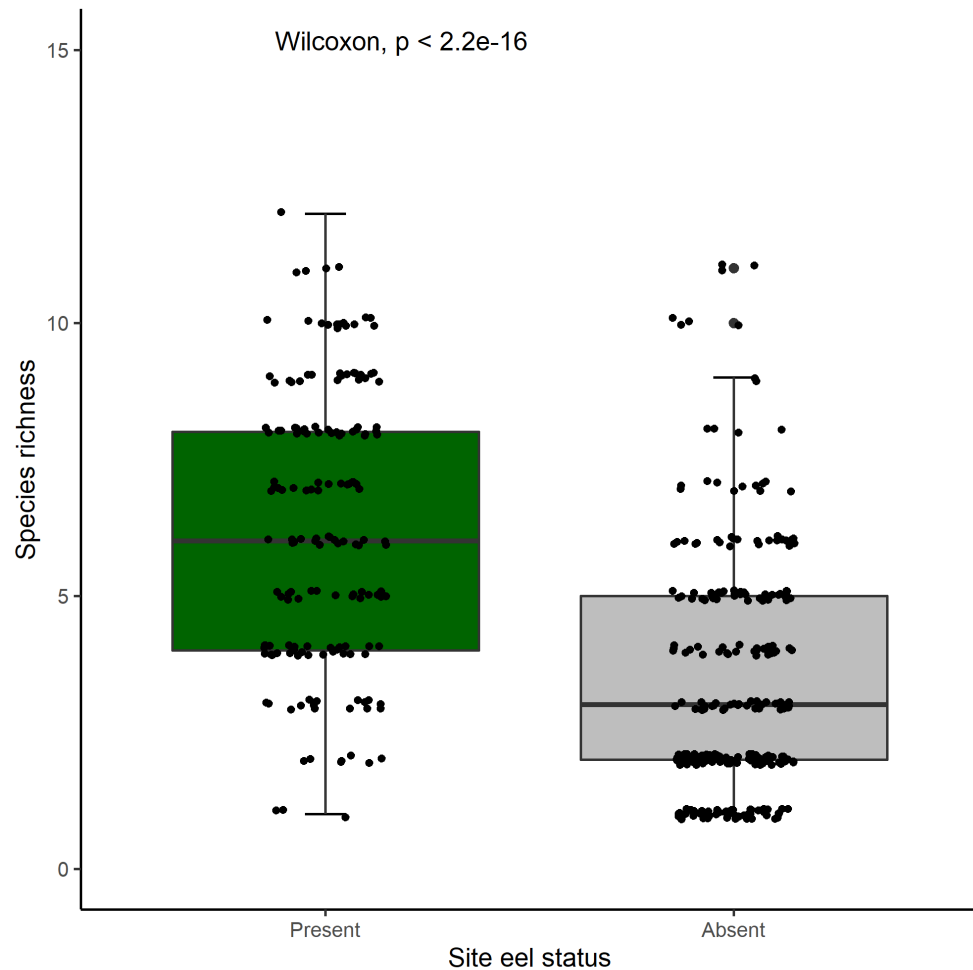
>95% confidence

>99% confidence

\*\*Numbers are rounded, 100% probability of absence does not exist!



# Site categorization



Jaccard index (PERMANOVA,  $R^2 = 0.08$ ,  $DF = 1$ ,  $P = 0.001$ )

Bray-Curtis (PERMANOVA,  $R^2 = 0.11$ ,  $DF = 1$ ,  $P = 0.001$ )

# Conclusions

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- We developed a model which allows assessment of the 'confidence in absence' of priority species.
- A >99% certainty that 27 of our sites were eel negative.
- **Can be applied to inform cost-benefit analysis and survey designs of future work (Confidence, Lab & Field resources).**

